

# LEAF WATER POTENTIALS OF SUNLIT AND/OR SHADED GRAPEVINE LEAVES ARE SENSITIVE ALTERNATIVES TO STEM WATER POTENTIAL

Larry E. WILLIAMS

Department of Viticulture and Enology  
University of California, Davis and Kearney Agricultural Center, 9240 S. Riverbend ave,  
Parlier, California 93648, United States

## Abstract

**Aims:** Leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials were measured on grapevines to determine the effects of shoot location on both methods to assess vine water status.

**Methods and results:** Cabernet-Sauvignon and Merlot used in this study were grown at two locations in California. Measurements were taken at midday in July (Merlot) and at two times of the day (morning and afternoon), on two dates in August (Cabernet-Sauvignon). Measurements of  $\Psi_l$  and  $\Psi_{stem}$ , stomatal conductance and transpiration were taken on shoots entirely exposed to direct solar radiation or on shoots totally in the shade at the times of measurement. There were significant differences ( $P < 0.05$ ) between  $\Psi_l$  and/or  $\Psi_{stem}$  measured on shoots exposed to direct solar radiation and those in the shade. Both  $\Psi_l$  and  $\Psi_{stem}$  were significantly greater on the shoots exposed to direct sunlight compared to those in the shade. There was no significant difference between  $\Psi_l$  measured on shaded leaves and  $\Psi_{stem}$  determined on the fully exposed shoots.

**Conclusions:** Regardless of method used, water potentials were highly correlated with stomatal conductance measured on leaves in direct sunlight at the same time. All means of measuring grapevine water potential used in this study were highly correlated with one another.

**Significance and impact of the study:** The data indicate that any of the techniques used in this study would be a sensitive indicator of vine water status and that the  $\Psi$  of shaded leaves would be an alternative to the measurement of  $\Psi_{stem}$ .

**Key words:** water potential, Cabernet-Sauvignon, Merlot

## Résumé

**Objectifs :** Le potentiel hydrique foliaire ( $\Psi_l$ ) et le potentiel tige ( $\Psi_{stem}$ ) ont été mesurés au niveau des vignes pour déterminer l'influence de la position des rameaux sur les deux mesures utilisées pour la détermination de l'état hydrique de la vigne.

**Méthodes et résultats :** Les cépages considérés dans cette étude sont le Cabernet-Sauvignon et le Merlot dans deux localités de Californie. Les mesures ont été réalisées à la mi-journée en juillet (Merlot) et deux fois pendant la journée (le matin et l'après-midi), à deux dates différentes en août (Cabernet-Sauvignon). Les mesures de  $\Psi_l$  et  $\Psi_{stem}$ , de la conductance stomatique et de la transpiration ont été faites au niveau des rameaux directement exposés au soleil ou des rameaux entièrement à l'ombre lors de la prise des mesures. Des différences significatives ( $P < 0.05$ ) ont été obtenues entre le  $\Psi_l$  et/ou le  $\Psi_{stem}$  mesurés sur les rameaux ensoleillés et ceux à l'ombre. Les deux potentiels ( $\Psi_l$  et  $\Psi_{stem}$ ) ont été significativement plus importants pour les rameaux ensoleillés par rapport à ceux à l'ombre. Cependant, aucune différence significative n'a été notée entre le  $\Psi_l$  des feuilles à l'ombre et le  $\Psi_{stem}$  des rameaux ensoleillés.

**Conclusions :** Indépendamment de la méthode utilisée, les mesures simultanées des potentiels hydriques et de la conductance stomatique des feuilles exposées au soleil ont présenté une forte corrélation. Tous les moyens de mesure du potentiel hydrique de la vigne considérés dans cette étude ont été fortement corrélés entre eux.

**Signification et impact de l'étude :** Les données obtenues indiquent que toutes les techniques utilisées dans cette étude sont considérées comme des indicateurs sensibles de l'état hydrique de la vigne et que le  $\Psi$  des feuilles à l'ombre pourrait être une alternative à la mesure du  $\Psi_{stem}$ .

**Mots clés :** potentiel hydrique, Cabernet-Sauvignon, Merlot

*manuscript received 25<sup>th</sup> July 2011 - revised manuscript received 5<sup>th</sup> March 2012*

## INTRODUCTION

Methods to assess the water status of plants used in mechanistic and drought tolerant studies and for use in irrigation management have been reviewed (Jones, 2004; 2007). Pre-dawn leaf water potential ( $\Psi_{PD}$ ) is often used in viticulture studies and critical values have been established for grapevines (Ojeda *et al.*, 2001). Pre-dawn  $\Psi$  has been shown to be correlated with stomatal conductance ( $g_s$ ) and leaf net  $CO_2$  assimilation rate ( $A$ ) (Schultz and Stoll, 2010; Williams and Araujo, 2002). However, the main disadvantages for the use of  $\Psi_{PD}$  are the time of its measurement (Schultz and Stoll, 2010) and the fact that it may come into equilibrium with the wettest portion of the soil profile (Ameglio *et al.*, 1999) which would limit its use in drip irrigated vineyards (Williams and Trout, 2005). Santesteban *et al.* (2011) also concluded that similar  $\Psi_{PD}$  values among vineyards did not indicate similar levels of water stress.

Mid-morning, midday or mid-afternoon leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials are also used by scientists conducting research on grapevines and in commercial vineyards around the world as an irrigation management tool. Stomatal conductance and/or  $A$  have been shown to be highly correlated with midday  $\Psi_l$  (Williams, 2012; Williams and Araujo, 2002) and  $\Psi_{stem}$  (Patakas *et al.*, 2005). Midday  $\Psi_l$  has been shown to be highly correlated with daily water use of mature, field-grown grapevines (Williams *et al.*, 2012) while Choné *et al.* (2001) concluded that  $\Psi_{stem}$  or the difference between  $\Psi_l$  and  $\Psi_{stem}$  provided an indirect measurement of leaf transpiration. Both midday  $\Psi_l$  and  $\Psi_{stem}$  have been shown to better reflect differences in soil water content, soil matric potential and applied water amounts than  $\Psi_{PD}$  under drip irrigation (Williams and Trout, 2005). Lastly, non-water stressed values of  $\Psi_l$  and  $\Psi_{stem}$  have been established for potential use in vineyard irrigation management (Williams and Baeza, 2007; Williams and Trout, 2005).

Stem  $\Psi$  is determined by covering the leaf blade with a plastic bag and a reflective material for a certain amount of time allowing the leaf's  $\Psi$  to come into equilibrium with that of the stem's  $\Psi$ . The time allowed for this equilibrium to occur on grapevines ranges from placing the bags on the leaves the day before the measurements (Salón *et al.*, 2005), 2 h (Romero *et al.*, 2010) to 1.5 h (Santesteban *et al.*, 2011) before measurements and at least 10 minutes before measurements are taken (Shackel, 2007). A more typical time of 1 h has been used by many (Choné *et al.*, 2001; De la Hera *et al.*, 2007; Marsal *et al.*, 2008; Olivo *et al.*, 2009; Patakas *et al.*, 2005; Williams and Araujo, 2002). Some feel that  $\Psi_{stem}$  has an advantage over  $\Psi_l$  since it is often assumed to better reflect the water status of the whole plant, is less affected by environmental

factors and is able to detect differences among treatments to a greater extent than  $\Psi_l$  (Choné *et al.*, 2001; Patakas *et al.*, 2005; van Leeuwen *et al.*, 2006), but this view is not shared by all (Intrigliolo and Castel, 2006; Santesteban *et al.*, 2011). It has been demonstrated that  $\Psi_{stem}$  is affected by vapor pressure deficit (VPD) at the time of measurement in trees (McCutchan and Shackel, 1992; Shackel *et al.*, 1997) and grapevines (Olivo *et al.*, 2009; Williams and Baeza, 2007), similar to the response of  $\Psi_l$  to VPD in grapevines (Williams and Baeza, 2007). Olivo *et al.* (2009) also reported that the response of  $\Psi_{stem}$  to VPD varied across the growing season. Numerous papers have reported that  $\Psi_l$  and  $\Psi_{stem}$  are highly correlated with one another (Salón *et al.*, 2005; Stevens *et al.*, 1995; Williams, 2010; Williams and Araujo, 2002). Therefore it is surprising others have reported that changes in  $\Psi_{stem}$  are not mimicked by changes in  $\Psi_l$  (Choné *et al.*, 2001; Patakas *et al.*, 2005). Such would likely occur if the technique used in those studies to measure  $\Psi_l$  did not involve enclosing the leaf blade in a plastic bag just prior to severing the petiole. The failure to do so would result in erroneous values of  $\Psi_l$  (Turner and Long, 1980; Williams and Araujo, 2002; Williams *et al.*, 2012).

Shackel (2007) has demonstrated that  $\Psi_{stem}$  could differ by 0.2 MPa on the same grapevine due to random chance and/or measurement error. The first objective of this study was to determine if  $\Psi_{stem}$  varied significantly on an individual grapevine. To accomplish this,  $\Psi_{stem}$  was determined on shoots entirely exposed to direct solar radiation or completely in the shade on the same vine at the time of measurement. It was felt that shoot transpiration would differ considerably when these two were compared and possibly result in differing values of  $\Psi_{stem}$ . In addition, the choice of these two shoot types reflects differences among studies (referred to in the previous paragraph) in selecting which leaf will be bagged to measure  $\Psi_{stem}$ .

It has been demonstrated that the  $\Psi$  of shaded leaves on grapevines follows the same diurnal pattern as that of leaves exposed to direct sunlight and respond to differences in the amount of water applied similarly to that of  $\Psi_l$  (van Zyl, 1987). It has also been shown that the  $\Psi$  of shaded leaves on almond (*Prunus domestica* L.) trees is highly correlated with  $\Psi_{stem}$  (Goldhamer and Fereres, 2001) as is the  $\Psi$  of shaded leaves on pistachio (*Pistacia vera* L.) trees with the crop water stress index (Testi *et al.*, 2008). Therefore the second objective of this study was to compare  $\Psi_l$  of shaded leaves to that of  $\Psi_{stem}$  measured on the two shoot types (sunlit and shaded) on the same vine. This was to determine whether shaded leaf  $\Psi$  was correlated with other means of assessing water status of grapevines and if it were a viable alternative to measuring either  $\Psi_l$  or  $\Psi_{stem}$ .

## MATERIALS AND METHODS

### 1. Vineyard sites

The first vineyard site was a *Vitis vinifera* L. cv. Merlot vineyard located near the city of Madera (lat. 36°55'N; long. 120°9'W) in the San Joaquin Valley of California. The vines were planted on their own roots with 2.13 and 3.66 m vine and row spacings, respectively. The vines were trained to a bilateral cordon. The trellis was a cordon wire at a height of 1.28 m and a foliage catch wire 0.3 m above that. The canopy that develops using this training/trellis system with no shoot positioning has typically been referred to as the 'California sprawl'. Vineyard rows were approximately east/west. Vines were drip-irrigated at 0.4, 0.8 or 1.2 of estimated evapotranspiration ( $ET_c$ ) once irrigation commenced. The three irrigation treatments were achieved using different numbers of emitters or emitters with different discharge rates. Vineyard  $ET_c$  was estimated as the product of reference ET ( $ET_o$ ) and seasonal crop coefficients ( $K_c$ ) (Allen *et al.*, 1998). The seasonal  $K_c$ s used to schedule irrigations at this site were developed by measuring the shade cast on the ground beneath the canopy at solar noon and then using the relationship between the percentage of shade and the  $K_c$  given in Williams and Ayars (2005). The shaded area beneath the canopy was determined with a digital camera as outlined in Williams and Ayars (2005). Reference ET was obtained from the California Irrigation Management Information System (CIMIS) weather station (#145) located ~15 km from the vineyard. Variables measured and calculations used to determine daily  $ET_o$  from CIMIS can be found in Snyder and Pruitt (1992). Irrigation treatments each year in this vineyard did not commence until midday  $\Psi_l$  reached -1.0 MPa for vines in the 1.2 irrigation treatment. Vines were irrigated once weekly, beginning on Friday and ending by Sunday, with applied water amounts equal to that required for the week. Applied water amounts were measured with inline (in the drip line) water meters.

The second vineyard site, located near Paso Robles (lat. 35°41'N; long. 120°39'W), was planted to *V. vinifera* cv. Cabernet-Sauvignon grafted onto the rootstock 5C. Vine and row spacings in the vineyard were 1.83 and 3.05 m, respectively. The vines were trained to bilateral cordons and the trellis was a VSP (vertical shoot positioning). Row direction was approximately north/south. The first irrigation treatment consisted of vines irrigated at 1.12 of estimated  $ET_c$ . The second treatment consisted of irrigating vines once every two weeks with 90.7 L (16.2 mm) of water per vine and was designated as the DD (dry down) treatment. The seasonal crop coefficients used were those for a VSP trellis at a row width of 3.05 m (Williams, 2010). Reference ET was obtained from the PR1 weather station operated by the Paso Robles Wine Country Alliance

(PRWCA), located ~3 km from the vineyard. Calculation of applied water amounts was similar to that described for the Merlot site. Vines were irrigated 1 to 3 times weekly, depending upon the required amounts.

### 2. Vine water status

Water potentials at both locations were measured as described by Williams and Araujo (2002). Specifically, leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials were measured with a pressure chamber (model 1000; PMS Instrument, Corvallis, OR) on fully expanded, mature leaves. Leaf blades for  $\Psi_l$  determinations were covered with a plastic bag, quickly sealed and petioles then cut within 1 to 2 sec. The time between leaf excision and chamber pressurization was generally less than 10 to 15 sec. Approximately 60 min before measurements, leaves for determination of  $\Psi_{stem}$  were enclosed in plastic bags covered with aluminum foil. The plastic bag and aluminum foil enclosing the leaf blades used for  $\Psi_{stem}$  measurements were also inserted into the chamber during pressurization. Stomatal conductance ( $g_s$ ) and leaf transpiration ( $E$ ) were measured with a steady-state diffusion porometer (model 1600; LI-COR, Lincoln, NE) on the leaves used for  $\Psi_l$  measurements.

Leaves selected for  $\Psi$  measurements were either from shoots entirely exposed to direct solar radiation (referred to as sunlit shoots) or shoots totally in the shade (referred to as shaded shoots) at the time of measurement. The leaves were generally located at node positions 7 or 8 from the base of the shoot. Leaves chosen for  $\Psi_l$  on the sunlit shoots were also exposed to direct sunlight. Leaves chosen for shaded  $\Psi_l$  were not exposed to any direct sunlight (such as sunflecks) prior to the time of measurement. Sunlit shoots were growing on the south side of the canopy while the shaded shoots were selected beneath shoots on the north side of the canopy in the Merlot vineyard. Measurements at this site were taken at midday (1300 – 1500 h Pacific daylight time [PDT]) on a single date. Measurements of  $\Psi$  and gas exchange were taken during the morning (0930 – 1030 h PDT) and afternoon (1530 – 1630 h PDT) on two different dates at the Paso Robles site. Sunlit and shaded shoots were selected from the east and west sides of the canopy, respectively, at this location during the morning measurement periods and just the opposite during the afternoon measurement periods. An additional sheet of aluminum foil was placed above the leaf chosen for  $\Psi_{stem}$  on the sunlit shoots at midday (Madera site) and during the afternoon (Paso Robles site) to further minimize heating effects. The sunlit and shaded shoots at both locations were growing on the same cordon close to one another but not necessarily on the same arm.

### 3. Temperature and relative humidity

Temperature and relative humidity were measured at both locations with two hand-held temperature/relative humidity probes (model DM-84 Multimeter with MultiMeterMateRH/T probe, A.W. Sperry Inst., Inc., Hauppauge, NY) and on occasions a Pocket Sling Psychrometer (Cole-Parmer, Vernon Hills, IL). The probes were positioned just beneath the canopy of vines trained to the VSP trellis (ensuring they were in the shade) and just below the fruiting zone of vines at the other vineyard site. The probes were placed at two different locations within two of the irrigation treatment plots. Measurements with the sling psychrometer were made between rows at a height of ~2 m. The probes were routinely calibrated in the laboratory and the outputs from the two were within 1°C and 2 % relative humidity. Photon flux density (PFD) was measured with a quantum sensor (model LI-190SA; LI-COR) or with the quantum sensor attached to the porometer when measuring shaded  $\Psi_l$  and  $g_s$ .

### 4. Data collection

Seven individual vines in each irrigation treatment at the Madera site were chosen for data collection. On each individual vine replicate a sunlit and shaded shoot were tagged and used for measurements.  $\Psi_l$  and  $\Psi_{stem}$  were determined on the same shoot of each shoot type (sunlit or shaded). At the Paso Robles site five and four individual vines within each irrigation treatment on the first and second data collection dates, respectively, were chosen and shoots tagged. The same vines were used for both the morning and afternoon measurements on the respective dates with other information as given for the Merlot study site. Data were analyzed via analysis of variance (ANOVA) using CoStat v. 6.400 (CoHort Software, Monterey, CA, USA). Means were separated using Duncan's Multiple range test. Linear correlations and regressions were also run on the data using CoStat.

**Table 1. Comparisons among leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials measured on shoots that were fully exposed to direct solar radiation (sunlit) or those that were entirely in the shade of the canopy (shaded) of Merlot grapevines growing near Madera in the San Joaquin Valley of California. Irrigation treatments included vines receiving applied water amounts at various fractions (0.4, 0.8 and 1.2) of estimated  $ET_c$ . Stomatal conductance ( $g_s$ ) and transpiration (E) values are also given for each treatment and shoot type.**

Irrigation Treatment (Fraction of $ET_c$ )	Sunlit Shoot		Shaded Shoot	
	( $\Psi_l$ )	( $\Psi_{stem}$ )	( $\Psi_l$ )	( $\Psi_{stem}$ )
$\Psi$ (MPa)				
1.2	-0.89 c	-0.59 b	-0.56 b	-0.46 a
0.8	-1.16 c	-0.92 b	-0.89 b	-0.80 a
0.4	-1.38 c	-1.18 b	-1.18 b	-1.06 a
$g_s$ (mmol m <sup>-2</sup> s <sup>-1</sup> )				
1.2	658 ± 15	--	285 ± 20	--
0,8	483 ± 26	--	194 ± 14	--
0,4	237 ± 21	--	98 ± 5	--
E (mmol m <sup>-2</sup> s <sup>-1</sup> )				
1.2	29.9 ± 0.7	--	12.4 ± 0.9	--
0,8	20.6 ± 1.1	--	7.7 ± 0.6	--
0,4	10.2 ± 0.9	--	3.9 ± 0.3	--

Measurements were taken on 25 July, 2002, between 1300 and 1500 hours. Ambient temperature, vapor pressure deficit (VPD) and photon flux density (PFD) during the measurement period averaged 36.2°C 4.84 kPa and 1758 mmol photons m<sup>-2</sup> s<sup>-1</sup>, respectively. Mean PFD for leaves measured in the shade were 157, 157 and 137 mmol photons m<sup>-2</sup> s<sup>-1</sup> for the 0.4, 0.8 and 1.2 irrigation treatments, respectively. Values of  $\Psi$  within a row for a particular irrigation treatment followed by a different letter are significantly different at  $P < 0.05$ . Values of  $g_s$  and E represent the means ± SE. (n = 7)

## RESULTS

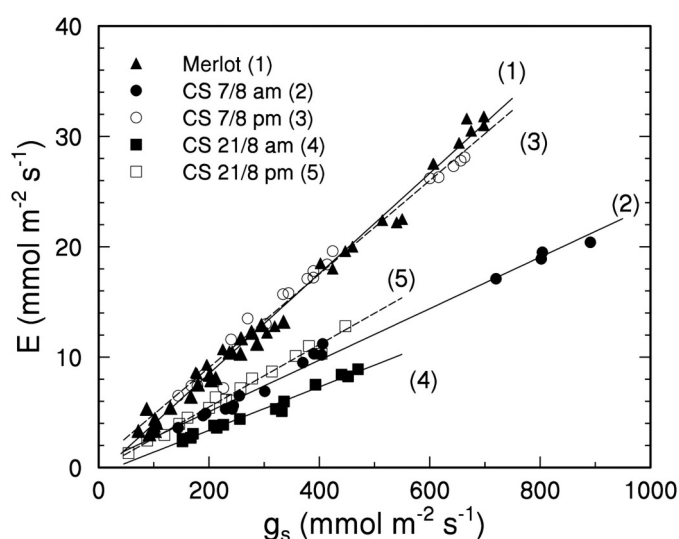
Estimated  $ET_c$  the week prior to 25 July at the Madera site was 30.8 mm (240 L vine<sup>-1</sup>). Applied water amounts the weekend prior to 25 July were 1.19, 0.79 and 0.42 of estimated  $ET_c$  for the 1.2, 0.8 and 0.4 irrigation treatments, respectively. Reference ET on 25 July was 6.6 mm at the Madera site.

This was the first year of the irrigation study in this Paso Robles vineyard and irrigation treatments were not imposed until 24 June. Estimated  $ET_c$  at the Paso Robles site for the week of 5 to 11 August was approximately 16.8 mm (~94 L vine<sup>-1</sup>) while that for the week of 20 to 26 August was approximately 15.5 mm (~87 L vine<sup>-1</sup>). Applied water amounts for the 1.12 treatment were 0.9 of estimated  $ET_c$  the week of 5 to 11 August while that for the week of 20 to 26 August was only 0.78 of estimated  $ET_c$ . As mentioned in the Materials and Methods section, the DD treatment was irrigated once every two weeks with ~90 L vine<sup>-1</sup>, which would be equivalent to ~50 % of estimated  $ET_c$  over a two week period at this time of the season. Vines in this treatment were irrigated 13 days prior to 7 August with ~80 L vine<sup>-1</sup> and then irrigated on 8 August with ~70 L vine<sup>-1</sup>. Therefore, vines of the DD

treatment had been irrigated 13 days prior to measurements taken on 21 August.

There were significant differences between  $\Psi_l$  measured on leaves from the sunlit and shaded shoots of Merlot grapevines across the irrigation treatments (Table 1). There were also significant differences in  $\Psi_{stem}$  between the two shoot types. However, there was no significant difference between  $\Psi_{stem}$  of the sunlit shoot and  $\Psi_l$  of the shaded shoot. There were significant differences among irrigation treatments regardless of the method used to measure vine water status. All four methods of measuring vine water status were equally sensitive ( $P < 0.001$  for all methods) in detecting those differences among treatments. Both  $g_s$  and  $E$  measured at midday decreased as applied water amounts decreased whether the measurements were taken on leaves in the sun or the shade.

Evaporative demand differed between dates and the two time-of-day measurement periods at Paso Robles. Ambient temperature and VPD the afternoon of 7 August was similar to those at the Madera site on 25 July. There were significant differences between  $\Psi_l$  measured on Cabernet-Sauvignon leaves of sunlit and shaded shoots regardless of irrigation treatment, time of day or date of measurement (Table 2). Similar results were obtained with the  $\Psi_{stem}$  data except on 21 August at 1000 h for the 1.12 irrigation treatment. There were no significant differences across treatments, dates or times between  $\Psi_{stem}$  of the sunlit shoot and  $\Psi_l$  of the leaves on the shaded shoots. There were significant differences in vine water status between the two irrigation treatments regardless of which technique was used across dates and time of day ( $P < 0.001$  for all comparisons). Values of  $g_s$  and  $E$  of Cabernet-Sauvignon were affected by irrigation treatment, shoot type (sunlit vs. shaded) and time of day (Table 3). The response of leaf  $E$  to  $g_s$  at the Paso Robles site was affected by time of day and the date the measurements were taken (Figure 1). The slope of that relationship the afternoon of 7 August was similar to that of Merlot. The VPD at the time of measurement in the Cabernet-Sauvignon vineyard the afternoon of 7 August and in the Merlot vineyard were 4.65 and 4.84 kPa, respectively. The VPDs at the time of measurements at Paso Robles the morning of 7 August and both times on 21 August were less than 2.6 kPa.



**Figure 1.** The relationship between leaf transpiration ( $E$ ) and stomatal conductance ( $g_s$ ) measured on Merlot and Cabernet-Sauvignon (CS) grapevines. Measurements on the Cabernet-Sauvignon vines took place on 7 and 21 August (7/8 and 21/8) either in the morning (am) or afternoon (pm). Data points include measurements taken on both sunlit and shaded leaves. Regressions for each data set are as follows: (1)  $y = -0.594 + 0.0454x$ ,  $R^2 = 0.99$ ; (2)  $y = 0.391 + 0.0233x$ ,  $R^2 = 0.99$ ; (3)  $y = 0.571 + 0.0424x$ ,  $R^2 = 0.98$ ; (4)  $y = -0.582 + 0.0197x$ ,  $R^2 = 0.98$ ; (5)  $y = -0.182 + 0.0283x$ ,  $R^2 = 0.99$ .

Both  $g_s$  and  $E$  (measured on sunlit and shaded leaves) were linearly related to  $\Psi_l$  (measured on sunlit and shaded leaves, respectively) at the Madera site (Figure 2). Correlations between  $g_s$  (measured on sunlit leaves) and  $\Psi_{stem}$  measured on sun and shade shoots and  $\Psi_l$  measured on shaded leaves were similar to that of  $\Psi_l$  with  $r$  values being 0.95, 0.93 and 0.94, respectively ( $P < 0.001$  for all correlations). Similar types of correlations with  $g_s$  data

**Table 2. Comparisons among leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials (MPa) measured on sunlit and shaded shoots of Cabernet-Sauvignon in a vineyard near Paso Robles, California. Irrigation treatments consisted of vines receiving applied water amounts at 1.12 of estimated  $ET_c$  (1.12) and vines that had not been irrigated for two weeks prior to measurement (DD). Ambient temperature and vapor pressure deficit (VPD) at the time of measurement are also given.<sup>a</sup>**

Date (day/mo)	Time <sup>b</sup> of Day (h)	Temp. (°C)	VPD (kPa)	Irrigation treatment	Shoot Type			
					Sunlit		Shaded	
					( $\Psi_l$ )	( $\Psi_{stem}$ )	( $\Psi_l$ )	( $\Psi_{stem}$ )
7/8	1000	25,2	2,19	1,12	-0.71 c	-0.52 b	-0.51 b	-0.46 a
				DD	-1.16 c	-0.93 b	-0.91 b	-0.83 a
	1600	34,1	4,65	1,12	-1.02 c	-0.83 b	-0.83 b	-0.68 a
				DD	-1.40 c	-1.21 b	-1.18 b	-1.08 a
21/8	1000	22,6	1,53	1,12	-0.56 c	-0.37 ab	-0.39 b	-0.35 a
				DD	-1.25 c	-0.93 b	-0.92 b	-0.78 a
	1600	28,4	2,64	1,12	-0.92 c	-0.68 b	-0.68 b	-0.53 a
				DD	-1.30 c	-1.05 b	-1.08 b	-0.91 a

<sup>a</sup> Values of  $\Psi$  within a row for a particular irrigation treatment are significantly different at  $P < 0.05$ . (n = 5 on 7 August and 4 on 21 August)

<sup>b</sup> The PFD on 7/8 and 21/8 at 1000 and 1600 hours were 1836 and 1825 and 1501 and 1541  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ , respectively

**Table 3. The effect of date, time of day, irrigation treatment and shoot type on leaf stomatal conductance ( $g_s$ ) and transpiration (E) measured on Cabernet-Sauvignon grapevines. Other information is as given in Tables 1 and 2.**

Date (day/mo)	Time of Day (h)	Irrigation treatment	Shoot Type			
			Sunlit	Shaded	Sunlit	Shaded
			(g <sub>s</sub> )		(E)	
			(mmol m <sup>-2</sup> s <sup>-1</sup> )			
7/8 <sup>a</sup>	1000	1,12	824 ± 70	254 ± 16	19.0 ± 0.7	5.8 ± 0.4
		DD	393 ± 8	196 ± 23	10.3 ± 0.4	4.9 ± 0.6
	1600	1,12	645 ± 14	360 ± 30	27.1 ± 0.5	14.4 ± 0.4
		DD	384 ± 27	182 ± 14	16.6 ± 0.2	7.4 ± 0.3
21/8 <sup>b</sup>	1000	1,12	439 ± 15	190 ± 17	8.3 ± 0.3	3.3 ± 0.3
		DD	311 ± 16	171 ± 15	5.2 ± 0.3	2.8 ± 0.3
	1600	1,12	398 ± 19	248 ± 14	11.1 ± 0.6	6.9 ± 0.4
		DD	216 ± 40	112 ± 25	5.8 ± 1.2	3.1 ± 0.8

<sup>a</sup> The PFD measured at the leaf blade surface for leaves on shaded shoots at 1000 and 1600 hours were 239 and 215  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ , respectively.

<sup>b</sup> The PFD measured at the leaf blade surface for leaves on shaded shoots at 1000 and 1600 hours were 175 and 186  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ , respectively.

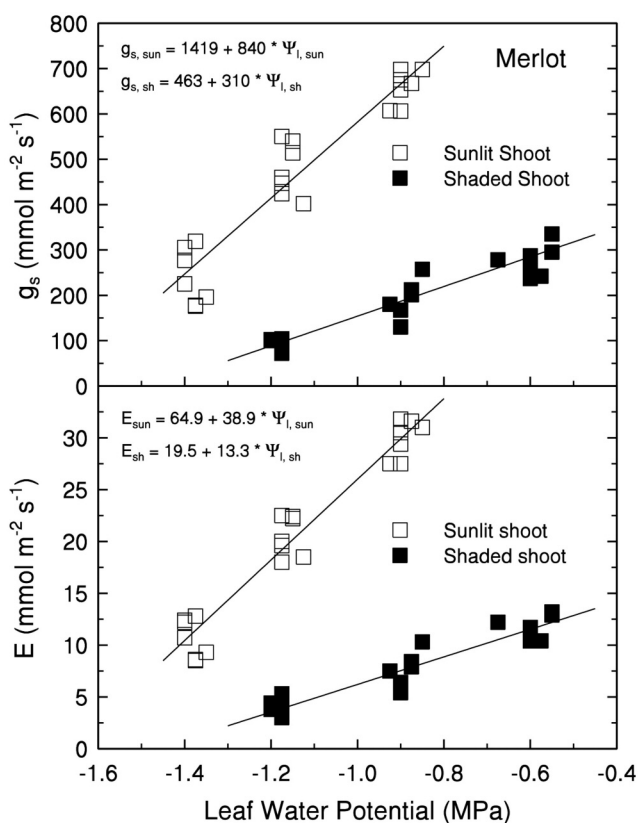
from the Paso Robles site were made but separating them as a function of time of day and date (Table 4). Stomatal conductance of sunlit leaves was significantly correlated with  $\Psi_l$  of sunlit leaves on both dates and times of day at Paso Robles (Table 4). Comparisons of  $g_s$  measured on sunlit leaves with  $\Psi_{stem}$  (sunlit shoot),  $\Psi_{stem}$  (shaded shoot) and  $\Psi_l$  (shaded leaves) were similar to that described for  $\Psi_l$  across dates and time of day. The relationships between

all measures of vine water status and E of sunlit leaves were comparable to those shown for  $g_s$ ;  $r$  and  $P$  values were similar.

There was a significant relationship between E (measured on sunlit leaves) and the difference between  $\Psi_l$  measured on sunlit leaves and  $\Psi_{stem}$  measured on shaded shoots ( $r = 0.73$ ,  $P < 0.001$ ) at the Madera site (data not given). There was also a significant relationship

between  $E$  (measured on sunlit leaves) and the difference between  $\Psi_l$  measured on sunlit leaves and  $\Psi_{stem}$  measured on shaded shoots the morning of 7 August ( $r = 0.90$ ,  $P < 0.01$ ) and in the afternoon on 21 August ( $r = 0.94$ ,  $P < 0.01$ ) at the Paso Robles site. However, there were no significant relationships between  $E$  and the difference between  $\Psi_l$  and  $\Psi_{stem}$  the afternoon of 7 August or the morning of 21 August at Paso Robles (data not given).

Morning and afternoon values of all techniques to measure vine water status at the Paso Robles site on both dates were significantly ( $P < 0.05$  on 7 August and  $< 0.001$  on 21 August) correlated with one another. The  $r$  values were  $> 0.76$  for all but the  $\Psi_{stem}$  (shaded shoots) comparison which was 0.65 on 7 August. The  $r$  values for all techniques to measure vine water status comparing afternoon with morning measurements on 21 August were in excess of 0.96 (this value being for the  $\Psi_{stem}$  of shaded shoots). Similar types of relationships were found between morning and afternoon measurements of  $g_s$  on sunlit leaves.

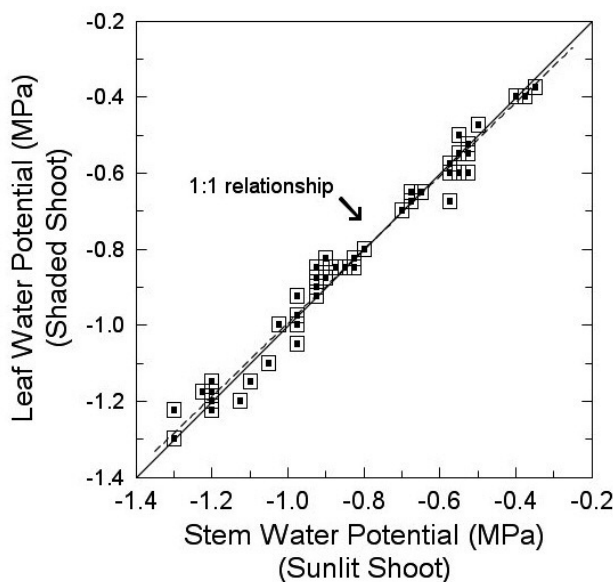


**Figure 2.** The relationships between stomatal conductance ( $g_s$ ) and leaf transpiration ( $E$ ) and leaf water potential ( $\Psi_l$ ) measured on Merlot grapevines. Stomatal conductance,  $E$  and  $\Psi_l$  data were measured either on sunlit or shaded shoots. The  $r$  values for the sun and shade  $g_s$  linear correlations were 0.95 and 0.94, respectively. The  $r$  values for the sun and shade  $E$  linear correlations were 0.97 and 0.95, respectively.

Comparisons were made between all techniques to measure vine water status across locations, time of day and dates (Table 5). All techniques were significantly correlated with one another. There was almost a 1:1 relationship between values of shaded  $\Psi_l$  and  $\Psi_{stem}$  of sunlit shoots (Figure 3).

## DISCUSSION

The data indicate that  $\Psi_{stem}$  can vary significantly on the same vine. The difference in  $\Psi_{stem}$  between the two shoot types of Merlot across irrigation treatments was fairly similar ( $\sim 0.1$  to  $0.12$  MPa). The differences in  $\Psi_{stem}$  between the two shoot types of Cabernet-Sauvignon ranged from none (the morning of 21 August for the 1.12 irrigation treatment) to  $0.15$  MPa. Shackel (2007) reported that any two values of  $\Psi_{stem}$  measured on the same grapevine differed by  $\pm 0.1$  MPa about 30 % of the time with differences of  $\pm 0.15$  MPa occurring 10 % of the time. The maximum difference between any two measurements reported by Shackel (2007) was  $\pm 0.2$  MPa. The maximum difference between any two  $\Psi_{stem}$  measurements on the same vine reported here was  $0.3$  MPa (1 out of 57 measurements) with a difference of  $0.2$  MPa the next highest value (2 out of 57 measurements). While the mean, maximum differences



**Figure 3.** The relationship between leaf water potential measured on shaded leaves and stem water potential measured on a sunlit shoot. Values represent data collected in the Merlot vineyard and across time of day and between dates in the Cabernet-Sauvignon vineyard.

The dotted line represents the linear correlation between these two parameters (see Table 5).

The solid line represents a 1:1 relationship.

**Table 4. The Pearson Product Moment Correlation Coefficient ( $r$ ) for the relationship between stomatal conductance ( $g_s$ ) measured on sunlit leaves and leaf ( $\Psi_l$ ) and stem ( $\Psi_{stem}$ ) water potentials of Cabernet-Sauvignon grapevines on two dates and two times of day in 2002. Other information is as given in Tables 2 and 3.**

Date (day/mo)	Time (h)	Shoot Type	Water Potential Method	$r$	Significance <sup>a</sup>
7/8	1000	Sun	$\Psi_l$	0,97	***
			$\Psi_{stem}$	0,97	***
		Shade	$\Psi_l$	0,98	***
			$\Psi_{stem}$	0,97	***
	1600	Sun	$\Psi_l$	0,95	***
			$\Psi_{stem}$	0,97	***
		Shade	$\Psi_l$	0,98	***
			$\Psi_{stem}$	0,97	***
21/8	1000	Sun	$\Psi_l$	0,93	***
			$\Psi_{stem}$	0,93	***
		Shade	$\Psi_l$	0,93	***
			$\Psi_{stem}$	0,93	***
	1600	Sun	$\Psi_l$	0,87	**
			$\Psi_{stem}$	0,82	*
		Shade	$\Psi_l$	0,82	*
			$\Psi_{stem}$	0,86	**

a \*, \*\* and \*\*\* represent significance at  $P < 0.05$ ,  $< 0.01$  and  $< 0.001$ , respectively.

reported in this study (0.15 MPa) between the two shoot types would appear to be minimal, measurements of vine water status in commercial vineyards are generally used to assign a ‘critical value’ after which an irrigation event would occur. The selection of a shoot located in the shade or the sun would provide differing values of  $\Psi_{stem}$  for the same vine.

The  $\Psi_{stem}$  of the sunlit shoot was always lower than that of the shaded shoot, never greater in this study. This was by design here as it was assumed transpiration of the sunlit shoot would be greater than that of the shaded shoot. The effect of an increase in  $E$  on  $\Psi_l$  due to increased frictional losses in the water pathway of plants can be described by rearranging the van den Honert (1948) equation (Jones, 1998):

$$\Psi_l = \Psi_{soil} - R_{sl} * E \quad (1)$$

where  $\Psi_{soil}$  is soil water potential and  $R_{sl}$  is the combined soil to leaf water flow resistance. The equation demonstrates that any change in  $E$  would affect  $\Psi_l$ . Transpiration of leaves on exposed shoots were 2 to almost 3 times greater than  $E$  of the shaded leaves on the same

vine with  $\Psi_l$  more negative for the sunlit leaves compared to the shaded leaves as would be predicted by Equation 1. One could also conclude that differences in shoot  $E$  would affect  $\Psi_{stem}$  for shoots on the same vine and that  $\Psi_{stem}$  of the shaded shoot would be less negative than that of the exposed shoot as was shown in this study.

All techniques used to assess vine water status ( $\Psi_l$ ,  $\Psi_{stem}$  and  $g_s$  on both sunlit and shaded shoots and leaves) in this study were highly correlated with one another. This would indicate that any of the techniques used would be a sensitive indicator of vine water status under the conditions of this study. The choice of which technique to use would be a function of the effect of the environment on the technique, reliability of the technique and its ease of use. It is often assumed that  $\Psi_l$  is more variable due to the effects of local environmental factors on  $E$  whereas  $\Psi_{stem}$  is less variable since the blades are covered and not transpiring (Choné *et al.*, 2001). However, Jones (1990) concluded that nearly all criticisms against the use of  $\Psi_l$  as a means to assess plant water status would be applicable to  $\Psi_{stem}$  to include that it is affected by environmental factors (Santesteban *et al.*, 2011). McCutchan and Shackel



**Table 5. Linear correlations among the various determinations of vine water status measured in this study. Data were generated from Merlot and Cabernet-Sauvignon grapevines (see Tables 1 and 2 for specific details). All equations were significant at the  $P < 0.001$  level. The term in the subscripts of the variables column following the comma, 'sun' and 'sh' represent shoot type, i.e. those that were exposed to direct sunlight or those that were entirely in the shade, respectively.**

**The Pearson Product Moment Correlation Coefficient ( $r$ ) is given in the rightmost column.  $n = 57$**

Variables	Equation	$r$
$\Psi_{l,\text{sun}}$ vs. $\Psi_{\text{stem},\text{sun}}$	$y = -0.285 + 0.943x$	0.972
$\Psi_{l,\text{sun}}$ vs. $\Psi_{l,\text{sh}}$	$y = -0.264 + 0.968x$	0.975
$\Psi_{l,\text{sun}}$ vs. $\Psi_{\text{stem},\text{sh}}$	$y = -0.353 + 0.991x$	0.956
$\Psi_{l,\text{sh}}$ vs. $\Psi_{\text{stem},\text{sun}}$	$y = -0.028 + 0.966x$	0.989
$\Psi_{l,\text{sh}}$ vs. $\Psi_{\text{stem},\text{sh}}$	$y = -0.086 + 1.031x$	0.987
$\Psi_{\text{stem},\text{sh}}$ vs. $\Psi_{\text{stem},\text{sun}}$	$y = 0.044 + 0.923x$	0.986

(1992) and Shackel *et al.* (1997) found that  $\Psi_{\text{stem}}$  of prune and deciduous nut and fruit trees, respectively, was responsive to VPD. Williams and Baeza (2007) found that  $\Psi_l$  and  $\Psi_{\text{stem}}$  of grapevines across cultivars and locations responded linearly to VPD, both decreased as VPD increased. Olivo *et al.* (2009) found that the response of a vine's  $\Psi_{\text{stem}}$  to VPD varied across the growing season but in general decreased as VPD increased. Both  $\Psi_l$  and  $\Psi_{\text{stem}}$  were also shown to be linearly reduced by increased ambient temperature (Williams and Baeza, 2007). Lastly, this study also demonstrated that the light environment of the shoot significantly affected  $\Psi_{\text{stem}}$ .

It is often assumed that  $\Psi_{\text{stem}}$  is able to detect differences in vine water status to a greater extent than that of  $\Psi_l$  and is more reliable (Choné *et al.*, 2001; Patakas *et al.*, 2005; van Leeuwen *et al.*, 2006). Values of  $\Psi_l$  and  $\Psi_{\text{stem}}$  in this study and those of Williams (2010), Williams and Araujo (2002) and Williams and Trout (2005) were significantly correlated with one another, as shown by others (Salón *et al.*, 2005; Stevens *et al.*, 1995). They were equally effective in detecting differences among irrigation treatments. Differences among studies concerning reliability and repeatability could be related to the method in which  $\Psi_l$  was measured. The technique used to measure  $\Psi_l$  in this study followed that described by Turner and Long (1980). The leaf blade was covered by a plastic bag just prior to severing the petiole and it remained on the blade through pressurization. This minimizes errors arising from rapid water loss subsequent to the blade being placed in the pressure chamber. Williams and Araujo (2002) and Williams *et al.* (2012) reported that differences in the  $\Psi$  of bagged leaves and those not bagged or only bagged after severing the petiole could result in differences of up to -0.6 MPa. Choné *et al.* (2001) did not bag leaves for the measurement of  $\Psi_l$  (X. Choné personal communication) while Patakas *et al.* (2005) only bagged the leaves after cutting the petiole (A. Patakas

personal communication). The techniques used in those two studies would have resulted in erroneous values, thereby affecting the reliability of  $\Psi_l$  in detecting differences among treatments.

A disadvantage of using  $\Psi_{\text{stem}}$  to determine plant water status is the fact that the leaf blade needs to be covered in order to stop E and allow the  $\Psi$  of the leaf to come into equilibrium with that of the stem. The time used for this equilibrium in many grapevine studies is on average 1 h (Choné *et al.*, 2001; De la Hera *et al.*, 2007; Marsal *et al.*, 2008; Olivo *et al.*, 2009; Patakas *et al.*, 2005; Williams and Araujo, 2002). This would require one to bag leaves in the vineyard and then return 1 h later to take measurements. Shackel (2007) reported that grapevine leaves in his study were bagged a minimum of 10 min to determine  $\Psi_{\text{stem}}$  on leaves in the lower canopy (it is assumed that those leaves were either in the shade or at least partially shaded). Fulton *et al.* (2001) had previously determined that an equilibrium period of 10 min or longer appeared suitable to provide an accurate  $\Psi_{\text{stem}}$  measurement on three different tree species (*Prunus dulcis*, *P. domestica* and *Juglans regia*). The minimal amount of time used in the Shackel (2007) and Fulton *et al.* (2001) studies to determine  $\Psi_{\text{stem}}$  indicates that the  $\Psi_l$  of the leaves chosen were close to that of  $\Psi_{\text{stem}}$ . In this study, the difference between  $\Psi_l$  of shaded leaves and  $\Psi_{\text{stem}}$  of the shaded shoots ranged from 0.04 to 0.15 MPa, not a large difference. Therefore, the selection of leaves in the shade may lessen the time required for equilibration and shorten the interval between bagging the leaves and measuring  $\Psi_{\text{stem}}$ . Unfortunately, the time required for the sunlit leaves'  $\Psi$ s to come into equilibrium with the stems'  $\Psi$ s were not determined in this study. It is also unknown whether a study such as that conducted by Fulton *et al.* (2001) has been conducted on grapevines.

Goldhamer and Fereres (2001) found that the  $\Psi$  of shaded almond leaves were highly correlated ( $R^2 = 0.94$ ) with  $\Psi_{\text{stem}}$  measured on the same trees. This only occurred once precautions were taken (shaded leaves were covered with a damp cloth prior to leaf excision) to minimize water loss during the measurement of shaded  $\Psi_1$ . In the Goldhamer and Fereres study, shaded  $\Psi_1$  was generally lower than that of  $\Psi_{\text{stem}}$ , similar to that found in this study for shaded  $\Psi_1$  compared to shaded shoot  $\Psi_{\text{stem}}$ . The high correlation between shaded  $\Psi_1$  and all other measures to assess vine water status in this study and the data of Goldhamer and Fereres (2001) and Testi *et al.* (2008) would indicate that shaded  $\Psi_1$  would be a viable alternative to the measurement of  $\Psi_{\text{stem}}$ . In addition, the almost 1:1 relationship between shaded  $\Psi_1$  and  $\Psi_{\text{stem}}$  of sunlit shoots would provide values of  $\Psi$  similar to  $\Psi_{\text{stem}}$  on a grapevine. Therefore the use of shaded  $\Psi_1$  to assess vine water status would not require an equilibrium period (bagging a leaf for a certain period of time to determine  $\Psi_{\text{stem}}$ ), thus saving time. The same could be said for the measurement of the  $\Psi$  of sunlit leaves. However, as with the measurement of  $\Psi_1$  precautions should be made to minimize water loss from the time the petiole is severed until the blade is placed in the pressure chamber to determine shaded  $\Psi_1$ .

Choné *et al.* (2001) reported that  $E$  was significantly correlated with  $\Psi_{\text{stem}}$  or the difference between  $\Psi_1$  and  $\Psi_{\text{stem}}$ , but not with  $\Psi_1$ . This differs from that found in this study where all measures of vine water status were significantly correlated with  $E$ . It was found here that the difference between  $\Psi_1$  of sunlit leaves and  $\Psi_{\text{stem}}$  of the shaded shoots (similar to the comparison in Choné *et al.*, 2001) was significantly correlated with  $E$  but the correlation coefficient (0.73) was much less than those for the comparisons of  $E$  (of sunlit leaves) and  $\Psi_1$  (of both sunlit and shaded leaves) and  $\Psi_{\text{stem}}$  (of both sunlit and shaded shoots). If the difference between  $\Psi_1$  and  $\Psi_{\text{stem}}$  were truly predictive of  $E$  then one would have found a significant correlation between shaded  $\Psi_1$  and shaded  $\Psi_{\text{stem}}$  and shaded leaf  $E$ . None were found in this study (data not given). Differences regarding the relationship of  $E$  and  $\Psi_1$  between this and the Choné *et al.* (2001) study is probably due to the method in which  $\Psi_1$  was measured in the two studies. Such differences have been noted earlier in the Discussion.

All measures of vine water status in this study were highly correlated with  $g_s$ . Shackel (2007) reported that  $g_s$  of Pinot noir was also highly correlated with both  $\Psi_1$  and  $\Psi_{\text{stem}}$  ( $R^2 = 0.88$  and  $0.85$ , respectively) as did Williams *et al.* (2012) for  $\Psi_1$ . The results from this study, those of Shackel (2007) and Williams *et al.* (2012) and that of De Bei *et al.* (2011), who stated that a good correlation exists between  $\Psi_1$  and  $g_s$  for many grapevine cultivars, are counter to the conclusions of Lovisolo *et al.* (2010), who

reported that there is no apparent relationship between midday measurements of grapevine  $g_s$  and  $\Psi_1$ .

The regulation of  $g_s$  in plants it thought to be controlled by hydraulic and/or non-hydraulic factors (Jones, 1998; Tardieu and Simonneau, 1998). The relationship between  $g_s$  and  $\Psi_1$  or shaded leaf  $g_s$  and shaded leaf  $\Psi_1$  in this and other studies on grapevines indicates possible hydraulic control of  $g_s$  with the direct mechanistic link being  $\Psi_1$ . However, Jones (1998) argued that the true driving variables for control of  $g_s$  are VPD and  $\Psi_{\text{soil}}$ . It has been demonstrated that grapevine  $\Psi_1$  and  $\Psi_{\text{stem}}$  are highly correlated with  $\Psi_{\text{soil}}$  (Williams and Trout, 2005). Therefore, the good relationship between  $g_s$  and all measures of vine water status in this study could be the result of the latter measurements being highly dependent upon  $\Psi_{\text{soil}}$ . In addition,  $g_s$  of vines in the 1.12 irrigation treatment at the Paso Robles site significantly decreased from morning to afternoon on 7 August, when VPD increased from 2.19 to 4.65 kPa, respectively, and corresponded to increases in  $E$  at a particular VPD (Figure 1). This is similar to the results of Cuevas *et al.* (2006), who found decreases in  $g_s$  between morning and afternoon measurements on Tempranillo grapevines planted to north/south rows. These results would indicate that VPD was also a driving variable of  $g_s$ . However, no such differences were found between  $g_s$  measurements in morning and afternoon for the DD treatment on 7 August or for the 1.12 treatment on 21 August. The small differences in VPD between morning and afternoon on 21 August may explain the lack of differences in  $g_s$  (sunlit leaves) of the 1.12 treatment vines whereas soil water availability for the DD treatment on 7 August would predominate with  $\Psi_{\text{soil}}$  controlling  $g_s$ .

Conversely,  $\Psi_1$  may be the dependent variable with  $g_s$ , via changes in  $E$ , the independent variable (Jones, 1998). The dependence of  $\Psi_1$  on  $E$  is also illustrated in this study. When leaf  $E$  increased from 12.4 (for the shaded leaves) to 29.9 mmol m<sup>-2</sup> s<sup>-1</sup> (for the sunlit leaves) of Merlot grapevines irrigated at 1.2 of estimated  $ET_c$  (Table 1),  $\Psi_1$  decreased from -0.56 to -0.89 MPa, respectively, (a change of 0.33 MPa) as one would predict from Equation 1. As leaf  $E$  increased, comparing leaves in the shade to those in the sun, by 12.9 (for the 0.8 treatment) and 6.3 mmol m<sup>-2</sup> s<sup>-1</sup> (for the 0.4 irrigation treatment),  $\Psi_1$  decreased by 0.27 and 0.2 MPa, respectively. Again the decreases in  $\Psi_1$  and their magnitude for these two treatments would be predicted from Equation 1. These results indicate that variations (to include diurnal changes) in  $\Psi_1$  and/or  $\Psi_{\text{stem}}$  on a single vine or vines within, for example an irrigation treatment, are the result of changes in  $g_s$  or  $E$  due to environmental factors (VPD and light). However, absolute values of  $\Psi_1$  ( $\Psi_{\text{stem}}$ ) and  $g_s$  are a function of the amount of water in the soil profile (or  $\Psi_{\text{soil}}$ ) once the soil is below field

capacity (Williams *et al.*, 2012). Thus, the influence of the environment on regulating  $\Psi_1$  via its effect on  $g_s$  and  $E$  diminishes as the soil dries and this has been demonstrated by Williams and Baeza (2007).

While all techniques used to measure vine water potential in this paper were sensitive indicators of vine water status and highly correlated with one another their relationships with vine growth and productivity were not assessed. However, the relationships between midday  $\Psi_1$  and daily whole vine  $E$  or various aspects of vine growth have been determined. It has been demonstrated that midday measurements of  $\Psi_1$  and  $g_s$  are highly correlated with the daily  $ET_c / ET_o$  ratio (Williams *et al.*, 2012) and that seasonal mean values of  $\Psi_1$  are highly correlated with vegetative and reproductive growth of Thompson Seedless (Grimes and Williams, 1990; Williams *et al.*, 2010a; 2010b) and Cabernet-Sauvignon grapevines (Williams, 2010). Based upon the results presented herein the other techniques used in this study to determine vine water status would more than likely also be correlated with those growth parameters and effectively used in an irrigation management program.

## CONCLUSIONS

The values of  $\Psi_1$  (ranging from -1.4 to -0.39 MPa) or  $\Psi_{stem}$  (ranging from -1.21 to -0.35 MPa) measured in this study provided a wide range of values in which conclusions could be drawn. All methods to assess vine water status, whether measuring  $\Psi_1$  or  $\Psi_{stem}$  on sunlit or shaded shoots (leaves) at mid-morning, midday or mid-afternoon, across cultivars and dates, were highly correlated with one another. They also reflected differences in the amount of water applied to vines in the various irrigation treatments. In addition,  $\Psi_1$ ,  $\Psi_{stem}$  and  $g_s$  measurements taken mid-morning at Paso Robles were highly correlated with those measurements taken during the afternoon. The above would indicate that any means to assess vine water status with the techniques utilized in this study would be an integrative indicator of whole vine water status under the conditions of this study. An important advantage of measuring  $\Psi$  on either sunlit or shaded leaves would be the increased numbers of samples one could take since it would not be necessary to bag the leaves from minutes to hours for equilibration as done with  $\Psi_{stem}$ . This would be most helpful where the measurement of vine water status is used as a tool in an irrigation management program. However, one would have to ensure that the measurement of  $\Psi_1$  included precautions to minimize water loss from the time the petiole was severed until pressurization within the chamber.

There are other factors which should be considered as a source of variation if any of the above techniques are

used in basic research or as a tool in an irrigation management program. The author in this study measured all variables, normalizing the data across locations, days and times. One may intuitively assume that  $\Psi_{stem}$  would provide a more reliable data set by minimizing operator error. However, Goldhamer and Fereres (2001) found that  $\Psi_{stem}$  values obtained by three different operators using the same trees varied significantly from one another throughout the day. Another source of significant variation in the use of plant based measures of vine water status would be vine to vine variability, even when  $\Psi_{stem}$  was utilized (van Leeuwen *et al.*, 2006). Such may have been minimized in this study as all treatments were irrigated on a regular basis and the vines' water status not entirely dependent upon soil water availability.

**Acknowledgements:** I would like to thank Steve Carter and Anji Perry of J. Lohr winery and John Simpson of Simpson Meadow Vineyards for their cooperation in this study. I would like to thank Dr. Ken Shackel for his helpful comments and Dr. A. Ezzahouani for the French translation of the abstract. This research was funded in part by the American Vineyard Foundation, Viticulture Consortium and the California Competitive Grants Program for Research in Viticulture and Enology.

## REFERENCES

- Allen R.G., Pereira L.S., Raes D. and Smith M. 1998. Crop evapotranspiration: guidelines for computing crop water requirements. Food and Agriculture Organization of the United Nations (FAO) Irrigation and drainage paper 56, Rome (Italy).
- Ameglio T., Archer P., Cohen M., Valancogne C., Daudet F.A., Dayau S. and Cruiziat P. 1999. Significance and limits in the use of predawn leaf water potential for tree irrigation. *Plant and Soil* **207**, 155-167.
- Choné X., van Leeuwen C., Dubourdieu D. and Gaudillere J.P. 2001. Stem water potential is a sensitive indicator of grapevine water status. *Annals of Botany* **87**, 477-483.
- Cuevas E., Baeza P. and Lissarrague J.R. 2006. Variation in stomatal behaviour and gas exchange between mid-morning and mid-afternoon of north-south oriented grapevines (*Vitis vinifera* L. cv. Tempranillo) at different levels of soil water availability. *Scientia Horticulturae* **108**, 173-180.
- De Bei R., Cozzolino D., Sullivan W., Cynkar W., Fuentes S., Damberg R., Pech J. and Tyermann S. 2011. Non-destructive measurement of grapevine water potential using near infrared spectroscopy. *Australian Journal of Grape and Wine Research* **17**, 62-71.
- De la Hera M.L., Romero P., Gómez-Plaza E. and Martínez A. 2007. Is partial root-zone drying an effective irrigation technique to improve water use efficiency and fruit quality in field-grown wine grapes under semiarid conditions? *Agricultural Water Management* **87**, 261-274.

- Fulton A., Buchner R., Olson B., Schwankl L., Gilles C., Bertagna N., Walton J. and Shackel K. 2001. Rapid equilibration of leaf and stem water potential under field conditions in almonds, walnuts and prunes. *HortTechnology* **11**, 609-615.
- Goldhamer D.A. and Fereres E. 2001. Simplified tree water status measurements can aid almond irrigation. *California Agriculture* **55**, 32-37.
- Grimes D.W. and Williams L.E. 1990. Irrigation effects on plant water relations and productivity of 'Thompson Seedless' grapevines. *Crop Science* **30**, 255-260.
- Intrigliolo D.S. and Castel J.R. 2006. Vine and soil-based measures of water status in a Tempranillo vineyard. *Vitis* **45**, 157-163.
- Jones H.G. 2007. Monitoring plant and soil water status: established and novel methods revisited and their relevance to studies of drought tolerance. *Journal of Experimental Botany* **58**, 119-130.
- Jones H.G. 2004. Irrigation scheduling: advantages and pitfalls of plant-based methods. *Journal of Experimental Botany* **55**, 2427-2436.
- Jones H.G. 1998. Stomatal control of photosynthesis and transpiration. *Journal of Experimental Botany* **49**, 387-398.
- Jones H.G. 1990. Physiological aspects of the control of water status in horticultural crops. *HortScience* **25**, 19-26.
- Lovisolo C., Perrone I., Carra A., Ferrandino A., Flexas J., Medrano H. and Schubert A. 2010. Drought-induced changes in development and function of grapevine (*Vitis* spp.) organs and in their hydraulic and non-hydraulic interactions at the whole-plant level: a physiological and molecular update. *Functional Plant Biology* **37**, 98-116.
- Marsal J., Mata M., del Campo J., Argones A., Vallverdú X., Girona J. and Olivo N. 2008. Evaluation of partial root-zone drying for potential field use as a deficit irrigation technique in commercial vineyards according to two different pipeline layouts. *Irrigation Science* **26**, 347-356.
- McCutchan H. and Shackel K.A. 1992. Stem-water potential as a sensitive indicator of water stress in prune trees (*Prunus domestica* L. cv. French). *Journal of the American Society for Horticultural Science* **117**, 607-611.
- Ojeda H., Deloire A. and Carbonneau A. 2001. Influence of water deficits on grape berry growth. *Vitis* **40**, 141-145.
- Olivo N., Girona J. and Marsal J. 2009. Seasonal sensitivity of stem water potential to vapour pressure deficit in grapevine. *Irrigation Science* **27**, 175-182.
- Patakas A., Noitsakis B. and Chouzouri A. 2005. Optimization of irrigation water use in grapevines using the relationship between transpiration and plant water status. *Agriculture, Ecosystems and Environment* **106**, 253-259.
- Romero P., Fernandez-Fernandez J.I. and Martinez-Cutillas A. 2010. Physiological thresholds for efficient regulated deficit-irrigation management in winegrapes grown under semiarid conditions. *American Journal of Enology and Viticulture* **61**, 300-312.
- Salón J.L., Chirivella C. and Castel J.R. 2005. Response of cv. Bobal to timing of deficit irrigation in Requena, Spain: Water relations, yield and wine quality. *American Journal of Enology and Viticulture* **56**, 1-8.
- Santesteban L.G., Miranda C. and Royo J.B. 2011. Suitability of pre-dawn and stem water potential as indicators of vineyard water status in cv. Tempranillo. *Australian Journal of Grape and Wine Research* **17**, 43-51.
- Schultz H.R. and Stoll M. 2010. Some critical issues in environmental physiology of grapevines: future challenges and current limitations. *Australian Journal of Grape and Wine Research* **16**, 4-24.
- Shackel K.A. 2007. Water relations of woody perennial plant species. *Journal International des Sciences de la Vigne et du Vin* **41**, 121-129.
- Shackel K.A., Ahmadi H., Biasi W., et al. 1997. Plant water status as an index of irrigation need in deciduous fruit trees. *HortTechnology* **7**, 23-29.
- Stevens R.M., Harvey G. and Aspinall D. 1995. Grapevine growth of shoots and fruit linearly correlate with water stress indices based on root-weighted soil matric potential. *Australian Journal of Grape and Wine Research* **1**, 58-66.
- Snyder R.L. and Pruitt W.O. 1992. Evapotranspiration data management in California. In: *Proceedings of the Irrigation and drainage sessions at Water Forum 1992*, EE, HY, IR, WR Div/ASCE, Baltimore, Md. USA.
- Tardieu F. and Simonneau T. 1998. Variability among species of stomatal control under fluctuating soil water status and evaporative demand: modelling isohydric and anisohydric behaviours. *Journal of Experimental Botany* **49**, 419-432.
- Testi L., Goldhamer D.A., Iniesta F. and Salinas M. 2008. Crop water stress index is a sensitive water stress indicator in pistachio trees. *Irrigation Science* **26**, 395-405.
- Turner N.C. and Long M.J. 1980. Errors arising from rapid water loss in the measurement of leaf water potential by the pressure chamber technique. *Australian Journal of Plant Physiology* **7**, 527-537.
- van den Honert T.H. 1948. Water transport in plants as a catenary process. *Discussions of the Faraday Society* **3**, 146-153.
- van Leeuwen C., Goutouly J.P., Costa-Ferreira A.M., Azaïs C., Marguerit E., Roby J.P., Choné X., Germain C., Homayouni S. and Gaudillere J.P. 2006. Intra-block variations of vine water status in time and space. In: *Proceedings of the VIth International Terroir Congress*, 3-5 July, 2006, Bordeaux, ENITA de Bordeaux, pp. 64-69.
- van Zyl J.L. 1987. Diurnal variation in grapevine water stress as a function of changing soil water status and meteorological conditions. *South African Journal of Enology and Viticulture* **8**, 45-52.
- Williams L.E. 2012. Effects of applied water amounts at various fractions of evapotranspiration (ET<sub>c</sub>) on leaf gas exchange of Thompson Seedless grapevines. *Australian Journal of Grape and Wine Research* **18**, 100-108.
- Williams L.E., Baeza P. and Vaughn P. 2012. Midday measurements of leaf water potential and stomatal

- conductance are highly correlated with daily water use of Thompson Seedless grapevines. *Irrigation Science* **30**, 201-212.
- Williams L.E. 2010. Interaction of rootstock and applied water amounts at various fractions of estimated evapotranspiration ( $ET_c$ ) on productivity of Cabernet-Sauvignon. *Australian Journal of Grape and Wine Research* **16**, 434-444.
- Williams L.E., Grimes D.W. and Phene C.J. 2010a. The effects of applied water at various fractions of measured evapotranspiration on water relations and vegetative growth of Thompson Seedless grapevines. *Irrigation Science* **28**, 221-232.
- Williams L.E., Grimes D.W. and Phene C.J. 2010b. The effects of applied water at various fractions of measured evapotranspiration on reproductive growth and water productivity of Thompson Seedless grapevines. *Irrigation Science* **28**, 233-243.
- Williams L.E. and Baeza P. 2007. Relationships among ambient temperature and vapor pressure deficit and leaf and stem water potentials of fully irrigated, field-grown grapevines. *American Journal of Enology and Viticulture* **58**, 173-181.
- Williams L.E. and Ayars J.E. 2005. Grapevine water use and the crop coefficient are linear functions of the shaded area measured beneath the canopy. *Agricultural and Forest Meteorology* **132**, 201-211.
- Williams L.E. and Trout T.J. 2005. Relationships among vine- and soil-based measures of water status in a Thompson Seedless vineyard in response to high-frequency drip irrigation. *American Journal of Enology and Viticulture* **56**, 357-366.
- Williams L.E. and Araujo F. 2002. Correlations among predawn leaf, midday leaf, and midday stem water potential and their correlations with other measures of soil and plant water status in *Vitis vinifera*. *Journal of the American Society for Horticultural Science*, **127**, 448-454.